

**Last Menstrual Period:**

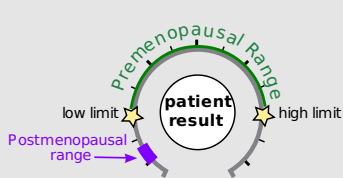
**Ordering Physician:**  
Precision Analytical

**DOB:** 1989-10-10  
**Age:** 26  
**Gender:** Female

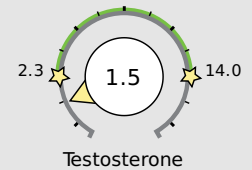
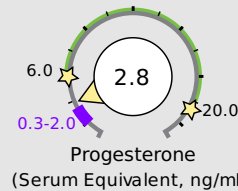
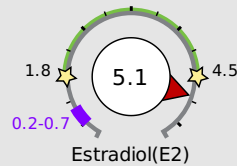
**Collection Times:**  
2016-10-02 06:00AM  
2016-10-02 08:00AM  
2016-10-01 06:00PM  
2016-10-01 10:00PM  
2016-10-02 02:00AM

## Hormone Testing Summary

### Key (how to read the results):

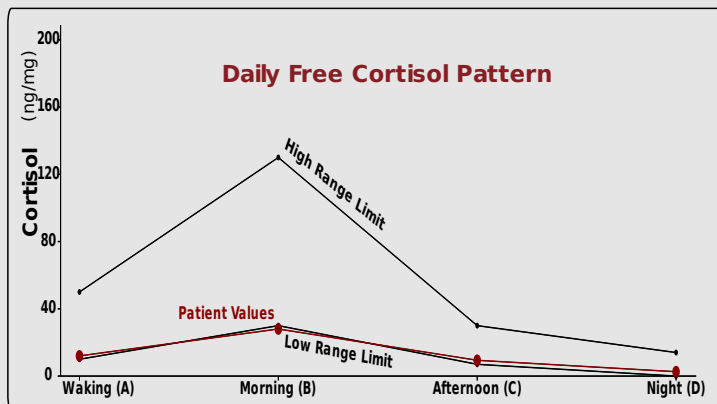


### Sex Hormones See Pages 2 and 3 for a thorough breakdown of sex hormone metabolites



Progesterone Serum Equivalent is a calculated value based on urine pregnanediol.

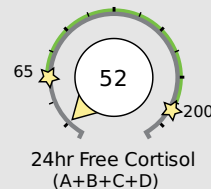
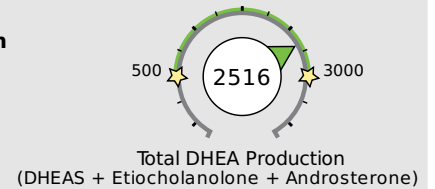
### Adrenal Hormones See pages 4 and 5 for a more complete breakdown of adrenal hormones



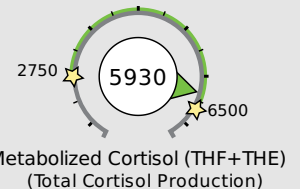
Free cortisol best reflects tissue levels. Metabolized cortisol best reflects total cortisol production.

### Total DHEA Production

Age	Range
20-39	1300-3000
40-60	750-2000
>60	500-1200



cortisol  
metabolism



The following videos (which can also be found on the website under the listed names along with others) may aid your understanding:

[DUTCH Complete Overview](#) [Estrogen Tutorial](#) [Female Androgen Tutorial](#) [Cortisol Tutorial](#)

**PLEASE BE SURE TO READ BELOW FOR ANY SPECIFIC LAB COMMENTS. More detailed comments can be found on page 9.**



**Accession # 00280399**

Female Sample Report  
123 A Street  
Somertown , CA 90266



**Sex Hormones and Metabolites**

**Last Menstrual Period:**

**Ordering Physician:**  
Precision Analytical

**DOB:** 1989-10-10  
**Age:** 26  
**Gender:** Female

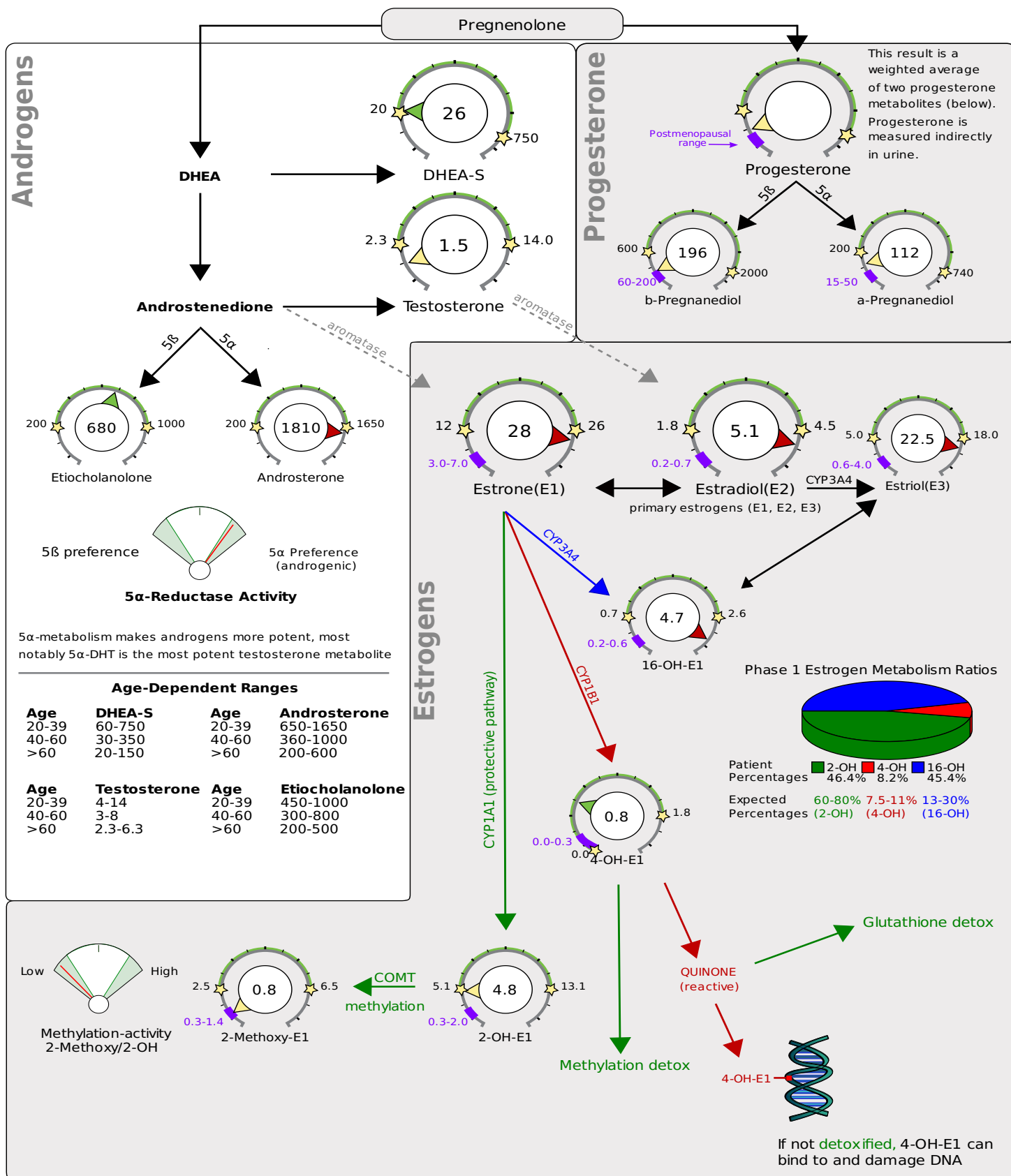
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Test	Result	Units	Luteal*	Postmenopausal	
<b>Progesterone Metabolites (Urine)</b>					
			<b>Range</b>	<b>Range</b>	
b-Pregnanediol	Below luteal range	196.0	ng/mg	600 - 2000	60-200
a-Pregnanediol	Below luteal range	112.0	ng/mg	200 - 740	15-50
<b>Estrogens and Metabolites (Urine)</b>					
Estrone(E1)	Above luteal range	28.2	ng/mg	12 - 26	3.0-7.0
Estradiol(E2)	Above luteal range	5.1	ng/mg	1.8 - 4.5	0.2-0.7
Estriol(E3)	Above luteal range	22.5	ng/mg	5 - 18	0.6-4.0
2-OH-E1	Below luteal range	4.8	ng/mg	5.1 - 13.1	0.3-2.0
4-OH-E1	Within luteal range	0.8	ng/mg	0 - 1.8	0-0.3
16-OH-E1	Above luteal range	4.7	ng/mg	0.7 - 2.6	0.2-0.6
2-Methoxy-E1	Below luteal range	0.8	ng/mg	2.5 - 6.5	0.3-1.4
2-OH-E2	Low end of luteal range	0.19	ng/mg	0 - 1.2	0-0.3
4-OH-E2	Within luteal range	0.2	ng/mg	0 - 0.5	0-0.1
2-Methoxy-E2	Within luteal range	0.3	ng/mg	0 - 0.7	0-0.4
Total Estrogen	High end of range	67.59	ng/mg	35 - 70	4.0-15
<b>Androgens and Metabolites (Urine)</b>					
DHEA-S	Low end of range	26.0	ng/mg	20 - 750	
Androsterone	Above range	1810.0	ng/mg	200 - 1650	
Etiocholanolone	Within range	680.0	ng/mg	200 - 1000	
Testosterone	Below range	1.5	ng/mg	2.3 - 14	
5a-DHT	Above range	7.2	ng/mg	0 - 6.6	
5a-Androstenediol	Above range	42.0	ng/mg	12 - 30	
5b-Androstenediol	Within range	32.0	ng/mg	20 - 75	
Epi-Testosterone	Within range	8.8	ng/mg	2.3 - 14	

\*the Luteal Range is the premenopausal range. When patients are taking oral progesterone this range for progesterone metabolites is not luteal and reflects the higher levels expected when patients take oral progesterone. This test is intended to be taken in the luteal phase of the menstrual cycle (days 19-22 of a 28 day cycle) for premenopausal women. The ranges in the table below may be used when samples are taken during the first few days (follicular) of the cycle, during ovulation (days 11-14) or when patients are on oral progesterone. See the following pages for age-dependent ranges for androgen metabolites.

Additional Normal Ranges	Follicular	Ovulatory	Oral Pg (100mg)
b-Pregnanediol	100-300	100-300	2000-9000
a-Pregnanediol	25-100	25-100	580-3000
Estrone (E1)	4.0-12.0	22-68	N/A
Estradiol (E2)	1.0-2.0	4.0-12.0	N/A

**Hormone metabolite results from the previous page are presented here as they are found in the steroid cascade. See the Provider Comments for more information on how to read the results.**





**Accession # 00280399**

Female Sample Report  
123 A Street  
Somertown , CA 90266



**Adrenal**

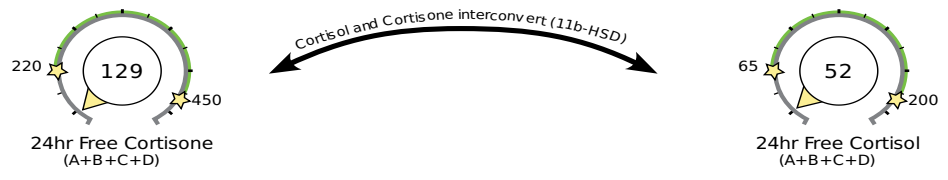
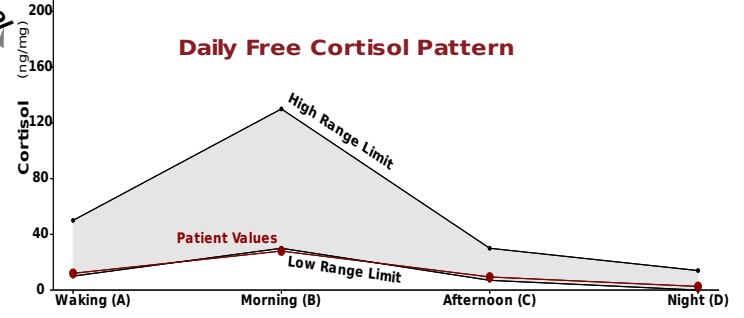
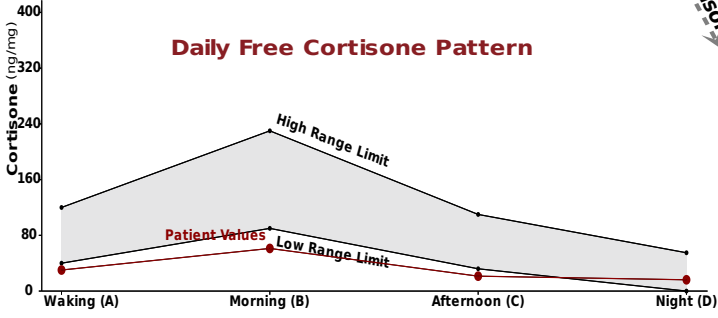
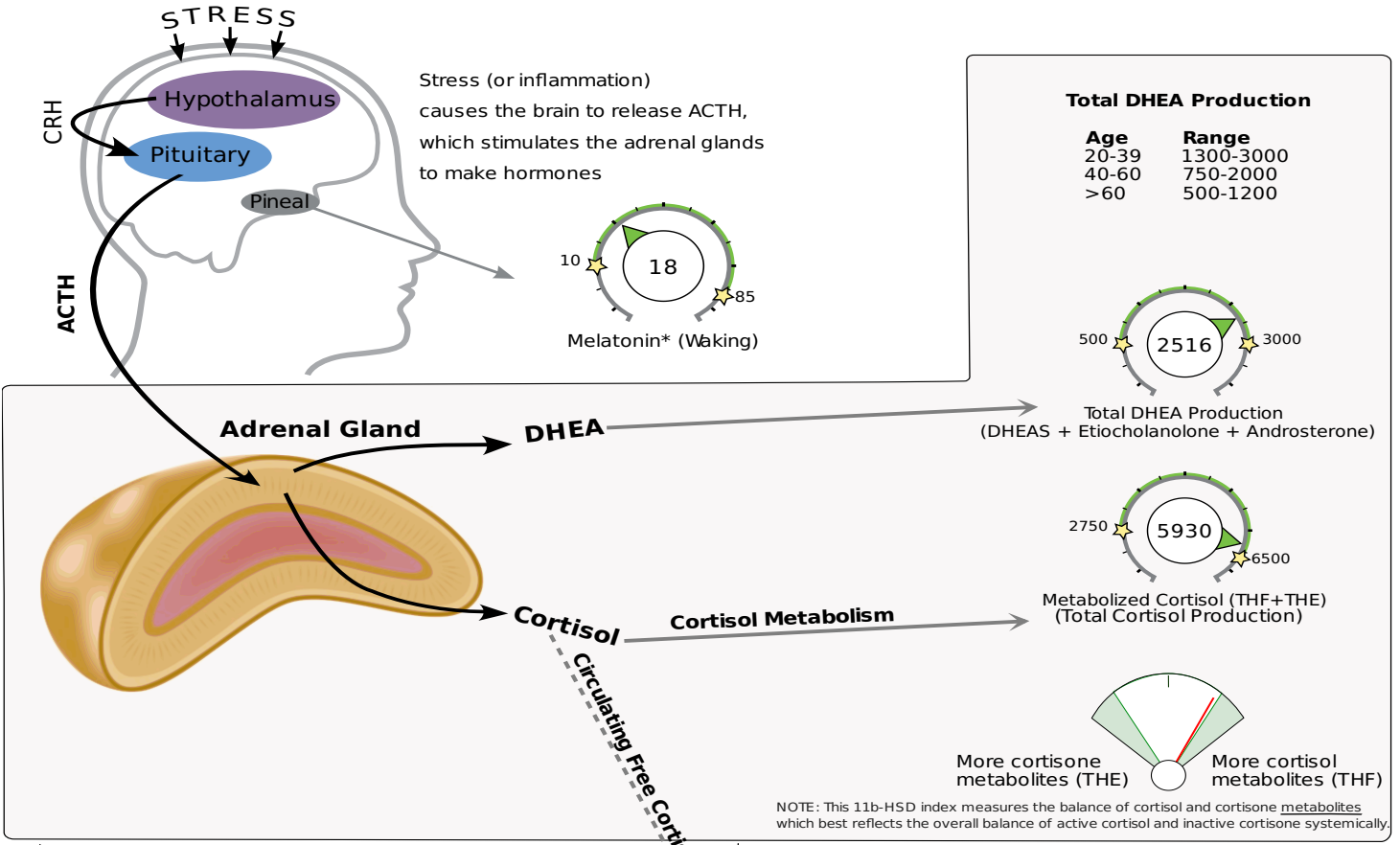
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2016-10-02 02:00AM

Category	Test		Result	Units	Normal Range
<b>Creatinine (Urine)</b>					
	Creatinine A (Waking)	Within range	0.4	mg/ml	0.2 - 2
	Creatinine B (Morning)	Within range	0.51	mg/ml	0.2 - 2
	Creatinine C (Afternoon)	Within range	0.92	mg/ml	0.2 - 2
	Creatinine D (Night)	Within range	1.01	mg/ml	0.2 - 2
<b>Daily Free Cortisol and Cortisone (Urine)</b>					
	Cortisol A (Waking)	Low end of range	12.0	ng/mg	10 - 50
	Cortisol B (Morning)	Below range	28.0	ng/mg	30 - 130
	Cortisol C (Afternoon)	Low end of range	9.4	ng/mg	7 - 30
	Cortisol D (Night)	Low end of range	2.6	ng/mg	0 - 14
	Cortisone A (Waking)	Below range	30.2	ng/mg	40 - 120
	Cortisone B (Morning)	Below range	61.2	ng/mg	90 - 230
	Cortisone C (Afternoon)	Below range	21.4	ng/mg	32 - 110
	Cortisone D (Night)	Within range	16.2	ng/mg	0 - 55
	24hr Free Cortisol	Below range	52.0	ng/mg	65 - 200
	24hr Free Cortisone	Below range	129.0	ng/mg	220 - 450
<b>Cortisol Metabolites and DHEA-S (Urine)</b>					
	a-Tetrahydrocortisol (a-THF)	Above range	400.0	ng/mg	75 - 370
	b-Tetrahydrocortisol (b-THF)	Above range	2500.0	ng/mg	1050 - 2500
	b-Tetrahydrocortisone (b-THE)	Within range	3030.0	ng/mg	1550 - 3800
	Metabolized Cortisol (THF+THE)	High end of range	5930.0	ng/mg	2750 - 6500
	DHEA-S	Low end of range	26.0	ng/mg	20 - 750



The first value reported (Waking "A") for cortisol is intended to represent the "overnight" period. When patients sleep through the night, they collect just one sample. In this case, the patient did not report waking up during the night to collect a sample, so the "Waking (A)" cortisol and cortisone values should accurately represent the entirety of the overnight period.



**Accession # 00280399**  
 Female Sample Report  
 123 A Street  
 Sometown, CA 90266



**Organic Acid Tests (OATs)**

**Last Menstrual Period:**

**Ordering Physician:**  
 Precision Analytical

**DOB:** 1989-10-10  
**Age:** 26  
**Gender:** Female

**Collection Times:**  
 2016-10-02 06:00AM  
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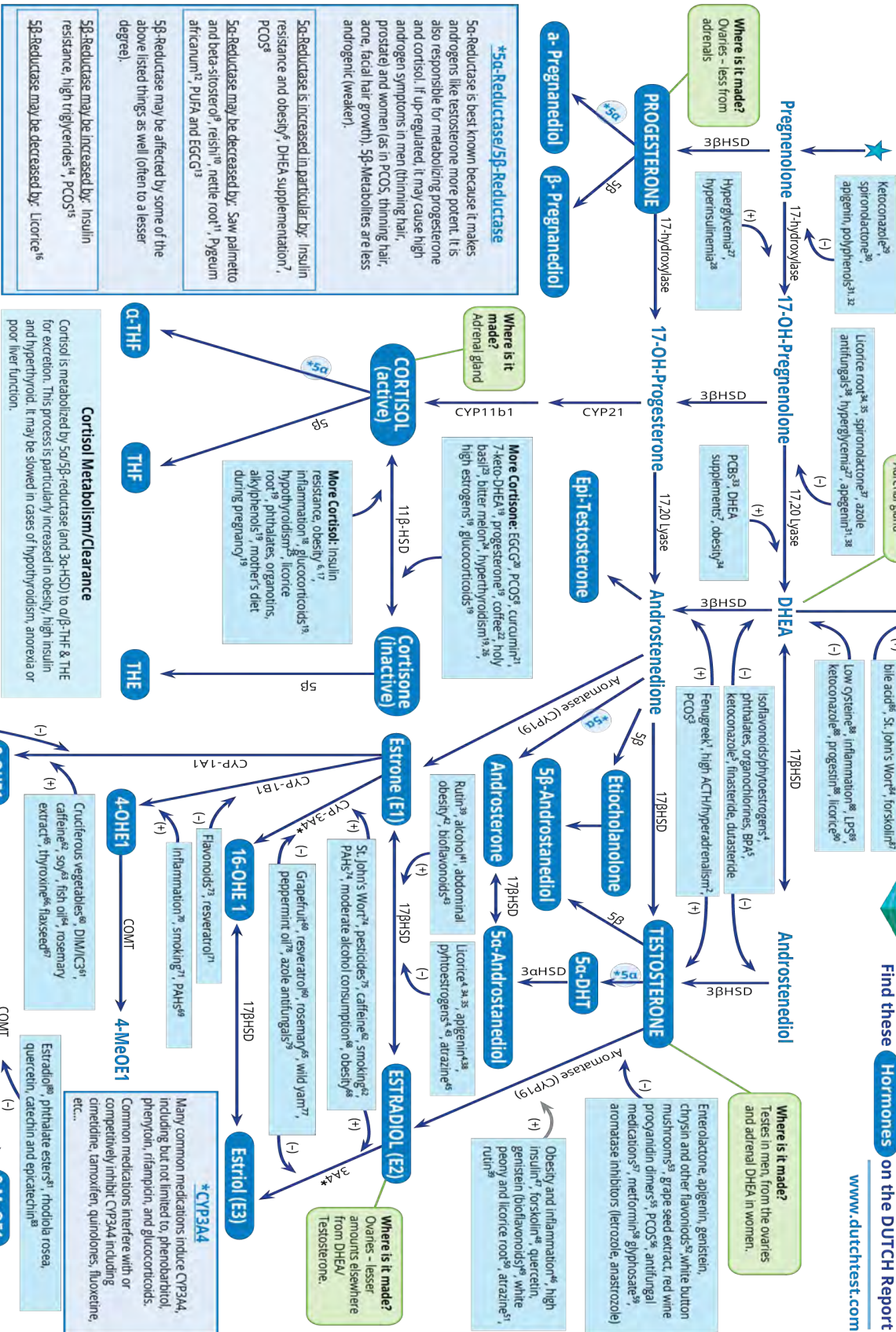
Category	Test	Result	Units	Normal Range
<b>Nutritional Organic Acids</b>				
Vitamin B12 Marker (may be deficient if high) - (Urine)				
	Methylmalonate (MMA)	Within range	1.2 ug/mg	0 - 2.2
Vitamin B6 Markers (may be deficient if high) - (Urine)				
	Xanthurenate	Above range	6.8 ug/mg	0 - 1.4
	Kynurenate	Above range	35.5 ug/mg	0 - 7.3
Glutathione Marker (may be deficient if low or high) - (Urine)				
	Pyroglutamate	Below range	23.2 ug/mg	32 - 60
<b>Neurotransmitter Metabolites</b>				
Dopamine Metabolite - (Urine)				
	Homovanillate (HVA)	Low end of range	5.6 ug/mg	4 - 13
Norepinephrine/Epinephrine Metabolite - (Urine)				
	Vanilmandelate (VMA)	Within range	4.8 ug/mg	2.4 - 6.4
Melatonin (*measured as 6-OH-Melatonin-Sulfate) - (Urine)				
	Melatonin* (Waking)	Low end of range	18.2 ng/mg	10 - 85
Oxidative Stress / DNA Damage, measured as 8-Hydroxy-2-deoxyguanosine (8-OHdG) - (Urine)				
	8-OHdG (Waking)	High end of range	4.3 ng/mg	0 - 5.2

Primary hormones (in CAPS) are made by organs by taking up cholesterol and converting it locally to, for example, progesterone. Much less is made from circulating precursors like pregnenolone. For example, taking DHEA can create testosterone and estrogen, but far less than is made by the testes or ovaries, respectively.

# Steroid Pathways

Find these Hormones on the DUTCH Report

www.dutchtest.com



20. Hirschgruber, J., et al., Green tea and one of its constituents, Epigallocatechin-3-gallate, are potent inhibitors of human 11 $\beta$ -hydroxysteroid dehydrogenase type 1. *PLoS One*, 2014, **9**(11): e114488.

21. Hu, G.X., et al., Curcumin as a potent and selective inhibitor of 11 $\beta$ -hydroxysteroid dehydrogenase-1: improving lipid profiles in high-fat-diet-treated rats. *PLoS One*, 2013, **8**(3): e64936.

22. Kanasou, A.G., et al., Coffee inhibits the reactivation of glucocorticoids by 11 $\beta$ -hydroxysteroid dehydrogenase type 1: a glucocorticoid coreceptor in the anti-diabetic action of coffee? *FEBS Lett*, 2006, **590**(17): p. 4031-5.

23. Jochle, R., et al., Ant-stress activity of Odrinum sanctum: Possible effects on Hypothalamo-pituitary-Adrenal Axis. *Phytother Res*, 2006, **20**(5): p. 805-14.

24. Blum, A., et al., Mometrolone diacetic acid: a novel 11 $\beta$ -hydroxysteroid type 2 diuretic, contains a specific 11 $\beta$ -hydroxysteroid dehydrogenase type 1 inhibitor. *J Steroid Biochem Mol Biol*, 2012, **128**(1-2): p. 51-5.

25. Hoshino, M., et al., Comprehensive study of urinary cortisone metabolites in hypertensive and hypotensive patients. *Clin Endocrinol (Oxf)*, 2006, **66**(4): p. 37-45.

26. Ishiyama, M., K. Horita, and Y. Imai, Urinary cortisone metabolites in the assessment of peripheral thyroid hormone action: application for diagnosis of resistance to thyroid hormone. *Thyroid*, 1993, **3**(3): p. 229-33.

27. Ueshima, H., et al., Decreased steroidogenic enzyme 17 $\alpha$ -HSD activity and increased 17 $\alpha$ -hydroxysteroid activities in type 2 diabetes mellitus. *Eur J Endocrinol*, 2002, **146**(3): p. 375-80.

28. Nestler, J.E. and D.J. Jawobenko, Decreases in ovarian cytochrome P450c17 alpha activity and serum free testosterone after reduction of insulin secretion in polycystic ovary syndrome. *N Engl J Med*, 1996, **335**(9): p. 617-23.

29. Engelhardt, D., et al., The influence of hexanocanone on human adrenal steroidogenesis: incorporation studies with tissue slices. *Clin Endocrinol (Oxf)*, 1991, **35**(2): p. 193-8.

30. Kossor, D.C. and H.D. Coody, Dose-dependent actions of spironolactone on the inner and outer zones of the guinea pig adrenal cortex. *Pharmacology*, 1992, **45**(1): p. 27-33.

31. Hasegawa, E., et al., Effect of polyphenols on the production of steroid hormones from human adrenocortical NCH-1295R cells. *Biol Pharm Bull*, 2013, **36**(2): p. 228-37.

32. Maril, N., et al., Bevacizumab inhibits androgen production of human adrenocortical h295R cells by lowering CYP17 and CYP21 expression and activities. *PLoS One*, 2012, **7**(12): p. e41742.

33. Andric, S.A., et al., Acute effects of polychlorinated biphenyl-concentrations and free testosterone fluids on rat testicular steroidogenesis. *Environ Health Perspect*, 2000, **108**(10): p. 955-9.

34. Kim, S.H., et al., Body fat mass is associated with Ratio of Steroid Metabolites Measured 17:20:spree Activity In Prepubertal Girls. *J Clin Endocrinol Metab*, 2016, **100**(12): p. 4653-4660.

35. Armanni, D., G. Bonanni, and M. Perrino, Regulation of serum testosterone in men by licochalcone. *Endocr Regul*, 1995, **44**(5): p. 115-8.

36. Armani, N., et al., Licochalcone serum testosterone in healthy women. *Steroid*, 2004, **69**(1-2): p. 703-6.

37. Serflin, P. and B.A. Lobo, The effects of spironolactone on adrenal steroidogenesis in hirsute women. *Fertil Steril*, 1985, **44**(5): p. 595-9.

38. Ayub, M. and M.J. Lovell, Inhibition of human adrenal steroidogenesis in vitro by imidazole drugs including ketoconazole. *J Steroid Biochem*, 1983, **32**(4): p. 515-24.

39. Wang, X., et al., Suppression of rat and human androgen biosynthesis enzymes by androgen: Possible use for the treatment of prostate cancer. *Endocrinology*, 2016, **111**: p. 66-72.

40. Hu, Y., et al., Brown adipose tissue activation by ucin analogates: polycystic ovary syndrome in rat. *Nat Biotechnol*, 2017, **47**: p. 21-28.

41. Serflin, T., et al., Acute effect of licochalcone on androgens in premenopausal women. *Aliment Pharmacol*, 2000, **35**(1): p. 84-90.

42. Corbould, A.M., et al., The effect of obesity on the ratio of type 3 17 $\alpha$ -hydroxysteroid dehydrogenase mRNA to cytochrome P450 aromatase mRNA in subcutaneous abdominal and intra-abdominal adipose tissue of women. *Int J Obes Relat Metab Disord*, 2002, **28**(2): p. 165-75.

43. Kazanietz, A., et al., Human 17 $\alpha$ -hydroxysteroid dehydrogenase type 3 is inhibited by dietary flavonoids. *Adv Exp Med Biol*, 2002, **505**: p. 151-61.

44. Le Bail, C., et al., Effects of phytoestrogens on aromatase, 3 $\beta$  and 17 $\beta$ -hydroxysteroid dehydrogenase activities and human breast cancer cells. *Life Sci*, 2000, **66**(14): p. 1281-91.

45. Ashrafian, S.O. and E.O. Farahni, Quercetin ameliorates atrazine-induced changes in the testicular function of rats. *Toxicol Ind Health*, 2016, **32**(7): p. 1278-85.

46. Gerard, C. and K.A. Brown, Obesity and breast cancer - Role of estrogen and the molecular underpinnings of aromatase regulation in breast adipose tissue. *Mol Cell Endocrinol*, 2018, **446**: p. 15-30.

47. Randolph, J.F., et al., The effect of fisalin on aromatase activity in isolated human endometrial glands and stroma. *Am J Obstet Gynecol*, 1987, **157**(6): p. 1534-9.

48. Watanabe, M. and S. Nakajin, Forskolin up-regulates aromatase (CYP19) activity and gene transcripts in the human endometrial carcinoma cell line h295R. *J Endocrinol*, 2004, **180**(1): p. 125-33.

49. Sanderson, J.T., et al., Induction and inhibition of aromatase (CYP19) activity by natural and synthetic flavonoid compounds in h295R human endometrial carcinoma cells. *Toxicol Sci*, 2004, **82**(1): p. 20-9.

50. Takeuchi, T., et al., Effect of paenolflavon, glycyrrhizin and glycyrrhonic acid on ovarian androgen production. *Am J Clin Med*, 1991, **19**(1): p. 33-8.

51. Holway, A.C., et al., Arazine-induced changes in aromatase activity in estrogen sensitive target tissues. *J Appl Toxicol*, 2006, **26**(8): p. 260-70.

52. Leprant, E.O., Modulation of Aromatase by Phytoestrogens. *Enzyme Res*, 2015, **2015**: p. 594655.

53. Novaes, M.R., et al., The effects of dietary supplementation with Agapaltes mushrooms and other medicinal fungi on breast cancer: evidence-based medicine. *Clinics (Sao Paulo)*, 2011, **66**(12): p. 2133-9.

54. Satoh, K., et al., Inhibition of aromatase activity by green tea extract catechins and their endocrinological effects of oral administration in rats. *Food Chem Toxicol*, 2002, **40**(7): p. 925-33.

55. Eng, E.T., et al., Suppression of estrogen biosynthesis by procyanidin dimer S in red wine and grape seeds. *Cancer Res*, 2003, **63**(23): p. 8316-22.

56. Chen, J., et al., The correlation of aromatase activity and obesity in women with or without polycystic ovary syndrome. *J Ovarian Res*, 2015, **8**: p. 11.

57. Ayub, M. and M.J. Lovell, The inhibition of human prostatic androgenase activity by imidazole drugs including ketoconazole and 4-hydroxyandrostenedione. *Drugs Pharmol*, 1990, **40**(7): p. 1569-75.

58. Rice, S.J., et al., Dual effect of metformin on growth inhibition and steroid production in breast cancer cells. *Int J Mol Med*, 2015, **35**(4): p. 1088-94.

59. Richards, S., et al., Differential effects of glyphosate and roundup on human placental cells and aromatase. *Environ Health Perspect*, 2005, **113**(6): p. 716-20.

60. Hodges, P.E. and D.M. Minich, Modulation of Metabolic Deoxy-Aromatic Pathways Using Foods and Food-Derived Compounds: A Scientific Review with Clinical Application. *Nutr Metab*, 2015, **2015**: p. 760689.

61. Michynova, J.J., H. Adlercreutz, and H.L. Bradlow, Changes in levels of microcytogen metabolites after oral intake of 3-carbonic flavonoids in humans. *J Natl Cancer Inst*, 1997, **89**(10): p. 718-23.

62. Somers, M.R., et al., Selected diet and lifestyle factors are associated with estrogen metabolism in a multicohort ethnic population of women. *J Nutr*, 2006, **136**(6): p. 1588-95.

63. Lu, L.L., et al., Increased urinary excretion of 2-hydroxyestrogen but not 16 $\alpha$ -hydroxyestrogen in premenopausal women during a soy diet containing isoflavones. *Cancer Res*, 2000, **60**(5): p. 1299-305.

64. Chen, H.M., et al., The combined effects of garlic oil and fish oil on the hepatic antioxidant and drug-metabolizing enzymes of rats. *Br J Nutr*, 2003, **89**(2): p. 183-200.

65. Deheer, P., et al., Induction of cytochrome P450 and/or detoxication enzymes by various extracts of rosemary: description of specific patterns. *Food Chem Toxicol*, 2001, **39**(9): p. 907-18.

66. Michynova, J.J. and S.A. Galbraith, Effects of exogenous thyroxine on C-62 and C-16 alpha hydroxylations of estradiol in humans. *Steroid*, 1990, **55**(1): p. 22-6.

67. Peters, L.P. and R.W. Teitel, Effect of high sucrose diet on cytochrome P450 1A and heterocyclic amine mutagenesis. *Anticancer Res*, 2003, **23**(14): p. 399-403.

68. Mahabir, S., et al., Effects of low-to-moderate alcohol supplementation on urinary estrogen metabolites in postmenopausal women in a controlled feeding study. *Cancer Med*, 2017, **6**(10): p. 2419-2423.

69. Uzes, B., et al., Resveratrol and its methoxy derivatives modulate the expression of estrogen metabolism enzymes in breast epithelial cells by Akt down-regulation. *Mol Cell Biochem*, 2017, **425**(1-2): p. 169-179.

70. Saito, M., et al., Up-regulation of CYP19B1 expression by anti-inflammatory glycosides is mediated by the p38 MAP kinase signal transduction pathway. *Carcinogenesis*, 2014, **35**(11): p. 2534-43.

71. Lu, M.X., et al., Estrogen receptor alpha promotes smoking carcinogen-induced lung carcinogenesis via cytochrome P450 1B1. *J Mol Med (Rev)*, 2015, **9**(31): p. 1221-33.

72. Jaramillo, L.C., et al., Effects of lipid components and combustion particle physicochemical properties on toxicological responses of lung cells. *J Environ Sci Health A Toxic Hazard Subst Environ Res*, 2018, **53**(4): p. 285-309.

73. Doostdar, H., M.D. Burke, and R.T. Mayer, Biochemical selective substrates and inhibitors for cytochrome P450 CYP19A and CYP17B1. *Toxicology*, 2000, **144**(3): p. 31-8.

74. Whitten, D.L., et al., The effect of St. John's wort extracts on CYP2A: a systematic review of prospective clinical trials. *Br J Clin Pharmacol*, 2006, **62**(5): p. 512-26.

75. Bredow, H.L., et al., Effects of pesticides on the ratio of 16 $\alpha$ -phar2-hydroxysterone a biologic marker of breast cancer risk. *Environ Health Perspect*, 1995, **103**(Suppl 7): p. 147-50.

76. Luckert, C., et al., Polycyclic aromatic hydrocarbons stimulate human CYP3A4 promoter activity via PKR. *Toxicol Lett*, 2013, **222**(2): p. 180-8.

77. Wu, W.H., et al., Estrogenic effect of yam ingestion in healthy postmenopausal women. *J Am Coll Nutr*, 2005, **24**(4): p. 235-43.

78. Dresser, G.K., et al., Evaluation of nempiprone oil and acetonyl palmitate as inhibitors of cytochrome P4503A4 activity in vitro and in vivo. *Clin Pharmacol Ther*, 2002, **72**(3): p. 247-55.

79. Nima, T., T. Inagawa, and H. Nishizaki, Drug interactions between nine anti-algal agents and drugs metabolized by human cytochrome P450. *Curr Drug Metab*, 2014, **15**(7): p. 651-75.

80. Jiang, H., et al., Human catechol-O-methyltransferase down-regulation by estradiol. *Neuropharmacology*, 2003, **45**(7): p. 1011-8.

81. Ho, P.W., et al., Effects of plasticizers and related compounds on the expression of the soluble form of catechol-O-methyltransferase in MCF-7 cells. *Curr Drug Metab*, 2008, **9**(4): p. 275-9.

82. Blum, K., et al., Manipulation of catechol-O-methyltransferase (COMT) activity to influence the attenuation of substance seeking behavior: a subtype of Reward Deficiency Syndrome (RDS). Is the pendulum upon gene polymorphisms a hypothesis. *Mol Hypotheses*, 2007, **6**(5): p. 1054-60.

83. van Duursen, M.B., et al., Phytochemicals inhibit catechol-O-methyltransferase activity in cytosolic fractions from healthy human mammary tissues: implications for catechol estrogen-mediated DNA damage. *Toxicol Sci*, 2004, **81**(2): p. 316-24.

84. Sehlin, A.O., et al., St. John's wort may ameliorate 2,4,6-trinitrobenzenesulfonic acid colitis off rats through the induction of pregnane X receptors and/or P-glycoproteins. *Physiol Pharmacol*, 2015, **66**(2): p. 203-14.

85. Pascuzzi, J.M., et al., Dexamethasone induces pregnane X receptor and retinoid X receptor-alpha expression in human hepatocytes: synergistic increase of CYP3A4 induction by pregnane X receptor activators. *Mol Pharmacol*, 2000, **58**(2): p. 361-72.

86. Zhou, H. and P.B. Hylton, Bile acids are nutrient signaling hormones. *Steroid*, 2014, **86**: p. 63-8.

87. Ding, X. and J.L. Stauffer, Induction of drug metabolism by forskolin: the role of the pregnane X receptor and the protein kinase A signal transduction pathway. *Pharmacol Exp Ther*, 2005, **312**(2): p. 849-56.

88. Mueller, J.W., et al., The Regulation of Steroid Action by Sulfation and Desulfation. *Endocr Rev*, 2015, **36**(5): p. 526-63.

89. Kim, M.S., et al., Suppression of DNA sulfotransferase (SULT2A1) during the acute phase response. *Am J Physiol Endocrinol Metab*, 2004, **287**(4): p. E731-8.

90. Al-Dajani, E.A., et al., Licochalcone and glycyrrhetic acid increase DNA and deoxyribonucleoside levels in vivo and in vitro by inhibiting adenosine SULT2A1 activity. *Mol Cell Endocrinol*, 2011, **336**(1-2): p. 102-9.

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# Clinical Support Overview

## Alert comments:

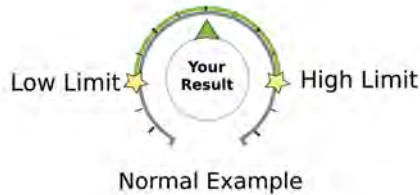
Thank you for choosing DUTCH for your functional endocrinology testing needs! We know you have many options to choose from when it comes to functional endocrinology evaluation, and we strive to offer the best value, the most up-to-date testing parameters and reference ranges, and the greatest clinical support to ensure the most accurate results.

Please take a moment to read through the Clinical Support Overview below. These comments are specific to the patient's lab results. They detail the most recent research pertaining to the hormone metabolites, treatment considerations, and follow-up recommendations. These comments are intended for educational purposes only. Specific treatment should be managed by a healthcare provider.

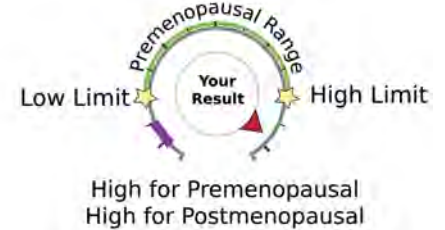
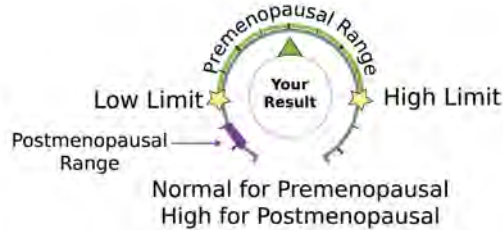
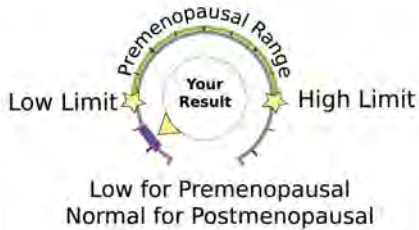
As an ordering provider, we encourage you to schedule a complimentary clinical consult with one of our expert DUTCH clinicians. Please schedule via our online scheduler, email [clinicalsupport@dutchtest.com](mailto:clinicalsupport@dutchtest.com) or call at 503.687.2050 to schedule a 15-minute or 30-minute consultation.

## How to read the DUTCH report

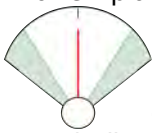
This report is not intended to treat, cure or diagnose any specific diseases. The graphic dutch dials in this report are intended for quick and easy evaluation of which hormones are out of range. Results below the left star are shaded yellow and are below range (left). Results between the stars and shaded green are within the reference range (middle). Results beyond the second star and shaded red are above the reference range (right). Some of these hormones also change with age, and the age-dependent ranges provided should also be considered.



For female reproductive hormones, a purple band is present on the dutch dials. This band represents the expected levels (reference range) for postmenopausal (or non-cycling) women.



In a few places on the graphical pages, you will see fan-style gauges. For sex hormones, you will see one for the balance between 5a/5b metabolism as well as methylation. For adrenal hormones, you will see one to represent the balance between cortisol and cortisone metabolites. These indexes simply look at the ratio of hormones for a preference. An average or "normal" ratio between the two metabolites (or groups of metabolites) will give a result in the middle (as shown here). If the ratio between the metabolites measured is "low" the gauge will lean to the left and similarly to the right if the ratio is higher than normal.



## Patient or Sample Comments

Throughout the provider comments you may find some comments specific to your situation or results. These comments will be found in this section or within another section as appropriate. Comments in other sections that are specific to your case will be in **bold**.

### The patient reports regular menstrual cycles.

Note: The dates listed on the samples imply that they were older than our allowed 3 weeks when they were received. The instructions ask that patients freeze or refrigerate samples if they are to be held. If that is not the case, the free cortisol and cortisone levels may drop somewhat over time if the samples are too old. Other hormones tested are stable for more than 12 weeks at room temperature. Samples that are refrigerated or frozen are stable for months.

## Progesterone Metabolism

Progesterone is made predominately in the ovaries by the corpus luteum following the release of an egg. Progesterone metabolite levels will increase to the premenopausal luteal range (the range established as the green band between the two gold stars) only after the release of an egg. The level of progesterone metabolites seen on the DUTCH test can help determine if ovulation occurred 5-7 days prior to test collection.

The primary role of progesterone is to prepare the endometrium of the uterus for implantation. In addition, it may balance the effects of estrogen, it is a neurosteroid, it acts as a diuretic and raises basal body temperature.

We are measuring metabolites of progesterone 5b-pregnanediol and 5a-pregnanediol. 5b-pregnanediol has less activity in the body but does represent a larger percent of total progesterone metabolism overall. 5a-pregnanediol is often a metabolite of more interest, as it can cross the blood brain barrier and up-regulate GABA activity and is considered neuroprotective to the brain. In some women the 5a-pregnanediol is also the cause of PMDD and irritability due to issues with the GABA receptor's inability to adjust for sensitivity to fluctuating

neurosteroids (Dr Briden).

If progesterone levels are in the low or lower end of the luteal reference range compared to estrogen levels, women may experience symptoms such as PMS, menorrhagia, mastalgia, moodiness, anxiety, and/or insomnia.

The metabolites of progesterone are excreted in urine (not the progesterone itself). When ordering the DUTCH Complete and DUTCH Plus reports, you will see a Progesterone Serum Equivalent on the summary page 1. The urine metabolites of progesterone have been proven to correlate strongly to serum progesterone. The Progesterone Serum Equivalent is most accurate with values in the luteal range and becomes more approximate at very low numbers in the postmenopausal range. Cycling women with very high progesterone metabolites may also decrease the accuracy of the serum equivalent calculation.

NOTE: If progesterone is taken orally (also with sublingual), these metabolites are elevated from gut metabolism and results do NOT accurately reflect serum levels.

**Progesterone metabolites are low for the luteal phase of the menstrual cycle. It is important to check in with the patient about the timing of the test in relation to menstruation before interpreting this result. If samples were collected too early or too late, the samples may not have been collected during the progesterone peak, which is reflected in the reference range. If the samples were collected about 5-7 days before menses, this likely indicates this cycle was anovulatory, indicating no fertile egg or progesterone production. Consider supporting the HPO axis (brain-to-ovary) communication and chasteberry extract, maca and B6 throughout the month to help with cycle regularity and ovulation.**

### Estrogen Metabolism

When evaluating estrogen levels, it is important to assess the following:

- **The status (low, normal or high?) of estrogen production:**

Levels of the primary ovarian product, estradiol (the strongest estrogen), as well as "total estrogens" may be considered. For women not on HRT, consider the appropriate range (premenopausal or postmenopausal).

- **Phase I Metabolism:**

Estrogen is metabolized (primarily by the liver) down three phase I pathways. The 2-OH pathway is considered the safest because of the anti-cancer properties of 2-OH metabolites. Conversely, the 4-OH pathway is considered the most genotoxic as its metabolites can create reactive products that damage DNA. The third pathway, 16-OH creates the most estrogenic of the metabolites (although still considerably less estrogenic than estradiol) - 16-OH-E1. If overall estrogen levels are high, production of 16-OH-E1 may exacerbate high estrogen symptoms. Similarly, a woman with very low levels of estrogens, may have less low estrogen symptoms if 16-OH metabolism is preferred. For example Armamento-Villareal showed that a higher 2-OH-E1/16-OH-E1 ratio correlated to bone loss (a low estrogen symptom). Estriol is thought of as a safer (weaker) estrogen metabolite, but it is important to remember that estriol is actually 16-OH-E2, so generally patients that make a lot of the potentially protective/weak estriol may also make a lot of the estrogenic 16-OH-E1.

When evaluating phase I metabolism, it may be important to look at the ratios of the three metabolites to see which pathways are preferred relative to one another. It may also be important to compare these metabolites to the levels of the parent hormones (E1, E2). If the ratios of the three metabolites are favorable but overall levels of metabolites are much lower than E1 and E2, this may imply sluggish phase I clearance of estrogens, which can contribute to high levels of E1 and E2. Similarly, patients with excessive phase I metabolism may have low E1 and E2 levels because of high rates of clearance (as opposed to simply not making a lot of estrogen).

The pie chart will assist you in comparing the three pathway options of phase I metabolism compared to what is "normal." 2-OH metabolism can be increased by using products containing D.I.M. or I-3-C. These compounds are found (or created from) in cruciferous vegetables and are known for promoting this pathway.

**Patients typically metabolize a much higher percentage of their estrogens down the more protective 2-OH pathway in phase 1 detoxification. Diindolylmethane (DIM) or Indole-3-Carbinol containing products can help move estrogens more efficiently down this pathway. Be aware that this typically lowers most of the other estrogens, including E1 and E2 as well. If the patients are taking or considering hormone replacement therapy, these products may be considered but a higher dose of estrogen may be needed for the same clinical effect if taken at the same time.**

- **Methylation (part of phase II metabolism) of estrogens:**

After phase I metabolism, both 4-OH and 2-OH (not 16-OH) estrogens can be deactivated and eliminated by methylation. The methylation-activity index shows the patient's ratio of 2-Methoxy-E1 / 2-OH-E1 compared to what is expected. Low methylation can be caused by low levels of nutrients needed for methylation and/or

genetic abnormalities (COMT, MTHFR). The COMT enzyme responsible for methylation requires magnesium and methyl donors. Deficiencies in folate or vitamin B6 or B12 can cause low levels of methyl donors. MTHFR genetic defects can make it more difficult for patients to make sufficient methyl donors. Genetic defects in COMT can make methylation poor even in the presence of adequate methyl donors.

## Androgen Metabolism

### Androgen Metabolites: DHEA

DHEA and androstenedione are made almost exclusively by the adrenal gland (although a smaller amount is made in the ovaries for). These hormones appear in urine as DHEA-S (DHEA-Sulfate), androsterone and etiocholanolone.

DHEA peaks for men and women in their 20's and 30's, with a slow decline expected with age. DHEA mainly circulates throughout the body as DHEA-s, with interconversion to active DHEA as it reaches various tissues. DHEA is a weak androgen and will predominately convert to androstenedione, which will then convert to testosterone or estrogen. DHEA-s is made by sulfation, has a much longer half-life than DHEA and largely lacks a diurnal rhythm, which is why it is considered the best way to assess DHEA levels in the body. DHEA-s levels can be affected both by the total production as well as by the body's ability to sulfate DHEA.

The best way to assess the total production of DHEA is to add up these three metabolites. As DHEA production decreases quite significantly with age, we provide the age-dependent ranges. Adrenal DHEA serves as the main source of estrogen, progesterone and testosterone for post-menopausal women.

**The Total DHEA Production (page 1) was about 2,516ng/mg which is within the overall range and also within the age-dependent range for this patient. This implies that the adrenal glands are producing appropriate DHEA levels.**

**The DHEA-S is lower than the other major metabolites of DHEA, etiocholanolone and androsterone. DHEA-S is mostly formed in the adrenal glands via sulfation. Inflammation can block sulfation. This lowers the DHEA-S and drives the 5a & 5b-reductase enzymes, metabolizing DHEA away from DHEA-S. Consider addressing inflammation, supporting sulfation with bile acid support (if needed), MSM, sulfur containing foods (such as arugula, asparagus, brassicas, onions, garlic, eggs) and molybdenum. Also consider supporting adrenal health through adaptogens and stress management.**

### • Androgen Metabolites: Testosterone

The DUTCH test measures the total of testosterone glucuronide and testosterone sulfate. These conjugates of testosterone are formed mostly from bioavailable testosterone that undergoes phase 2 metabolism to make it ready for urine excretion. Females make most of their DHEA in the adrenal gland and a fraction of that DHEA trickles down metabolically to testosterone. Testosterone is also made by the ovaries.

Testosterone glucuronide is mostly made by the UGT2B17 enzyme, which also makes the glucuronide forms of 5a-DHT and 5b-androstanediol. Genetic variants of this enzyme reduce the urinary levels of these hormones without affecting serum levels. The genetic variants of UGT2B17 vary in the population from 7-80% (variation dependent on genetic ancestry, with the highest rates in those of Asian descent). Heterozygous individuals show milder reductions in urinary testosterone than homozygous. For this reason, low and very low levels of urinary testosterone should be confirmed with serum testing before treatment is applied. Serum testing can include free and total testosterone and SHBG.

Testosterone levels may be better understood by also considering its downstream metabolites (5a-androstanediol, 5bandrostanediol). Technically, these metabolites can also be formed from DHEA metabolites without going through the testosterone pathway, but they generally tend to correlate with testosterone production.

Testosterone levels normally decline with age. Age dependent ranges are provided. Perimenopausal testosterone levels can transiently increase before declining again.

Androgens, specifically DHT and testosterone, help to support skin, connective tissue, bone and muscle integrity and promote dopamine conversion in the brain, which can help with mood and libido.

**The testosterone level for this patient is 1.50ng/mg, which is low. If the patient's symptoms do not fit with low testosterone, it is best to test serum levels to confirm before starting a treatment program due to the potential of falsely low urinary testosterone (see general notes for details). Androgens (DHEA and testosterone) in women help with muscle and weight**

**maintenance, memory and brain function, mood, libido and a sense of wellbeing. Lifestyle and diet modifications alone could be helpful to increase androgens such as weightlifting and high intensity interval training. Consider DHEA or TRT (testosterone replacement therapy), if appropriate and indicated. You could also consider using Tribulus, shatavari, maca, mitochondrial support and/or zinc to support androgens in women.**

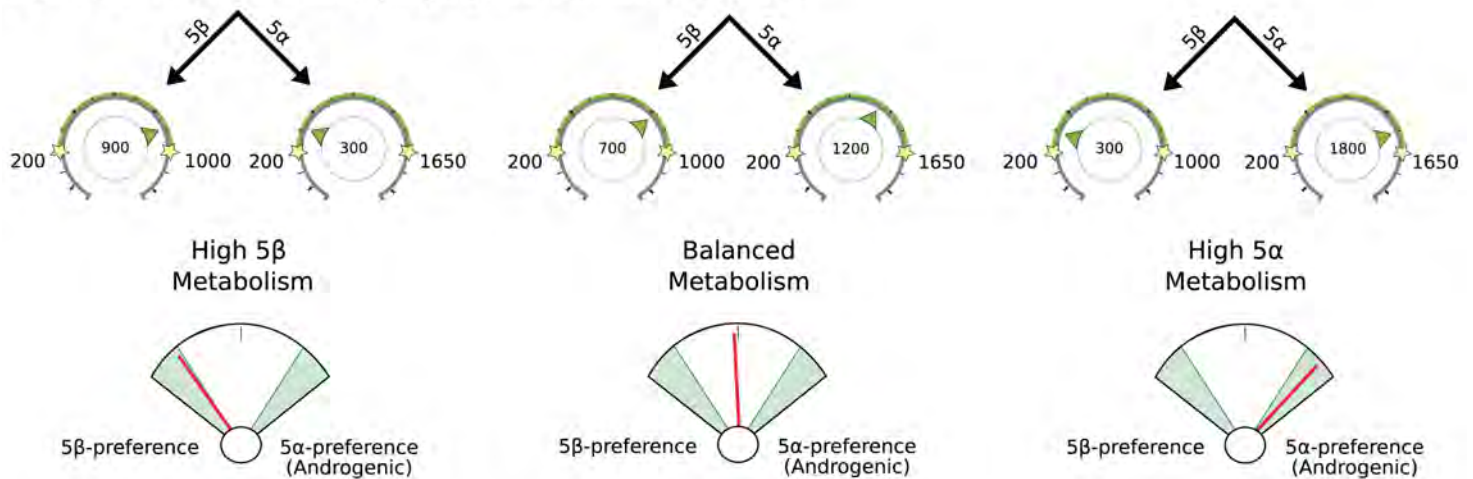
**• Androgen Metabolites: 5a-reductase versus 5b reductase**

5a-reductase converts testosterone into 5a-DHT (DHT), which is even more potent (~3x) than testosterone. High levels of DHT can lead to symptoms associated with too much testosterone, including scalp hair loss, hirsutism, acne and oily skin.

Metabolites created down the 5b-pathway are significantly less androgenic than their 5a counterparts.

The fan-style gauge below the hormones shows the 5a or 5b preference based on etiocholanolone (5b) and androsterone (5a) results. The gauge shows the relative ratio of 5a to 5b products but does not express the absolute value of DHT or if 5a-reductase inhibition is or is not indicated. Consider symptoms and look at the 5a-DHT result if high androgen symptoms are a concern. Progesterone metabolites are also metabolized by 5a and 5b enzymes and the balance between its two metabolites can be useful to confirm a 5a or 5b preference overall (or tissue specific preference).

Example of how to read fan-style gauge for 5a-reductase activity:



**While testosterone levels are not high, overall DHEA production is on the higher side and androgens are preferring the androgenic 5a pathway. Since the patient did not list significant symptoms of high androgens, these higher levels may be well tolerated by the patient. Since high insulin levels can lead to more DHEA production and 5a-metabolism, it may be worth exploring potential issues with blood sugar and/or insulin.**

When assessing androgens in women, it is important to consider DHEA and testosterone production, 5a-metabolism patterns as well as the patient symptoms. For example, a woman with higher levels of DHEA and testosterone will often have high androgen symptoms (facial hair, thinning scalp hair, etc.) exacerbated by 5a-metabolism.

If, on the other hand, she prefers 5b-metabolism she may not express high androgen symptoms in spite of higher levels of testosterone because 5b is the less androgenic pathway.

You will also see levels of epi-testosterone, which is not androgenic like testosterone. It happens to be produced in about the same concentrations as testosterone (this is an approximate relationship). This can be helpful when assessing the validity of urinary testosterone testing in an individual patient. If epi-testosterone is much higher than testosterone, serum testosterone assessment should be considered before initiating therapy for low testosterone. Epi-testosterone is suppressed when exogenous testosterone is given, which can serve as a proxy for assessing endogenous testosterone production which can be obscured by the exogenous hormone administration.

## DUTCH Adrenal

The HPA-Axis refers to the communication and interaction between the hypothalamus (H) and pituitary (P) in the brain down to the adrenal glands (A) that sit on top of your kidneys. When cortisol is needed in the body, the hypothalamus releases cortisol releasing hormone (CRH) and the pituitary responds by releasing adrenocorticotrophic releasing hormone (ACTH), which is the signal to the adrenal gland to release cortisol, DHEA and DHEA-s. It is these adrenal hormones that are assessed on the DUTCH test to understand the patient's HPA axis.

The cortisol awakening response (CAR) is a complex interaction between the HPA axis and the hippocampus, where ACTH normally surges right after waking leading to the day's highest levels of cortisol. The waking surge in cortisol helps with energy, focus, morning blood sugar and immune regulation.

As the day progresses, ACTH declines and subsequent cortisol decreases throughout the day, so it is low at night for sleep. This cycle starts over the next morning.

Free cortisol provides negative feedback to CRH & ACTH. When free cortisol is too low, ACTH will surge. ACTH will also surge when a physical or psychological stressor occurs.

Only a small fraction of cortisol is "free" and bioactive. The "free" cortisol is what the person feels in terms of energy and focus. Free cortisol is also what feeds back to the hypothalamus and pituitary gland for ACTH and cortisol regulation. The free cortisol daily pattern is very useful for understanding cortisol and its interaction with the patient's symptoms throughout the day. However, because only a fraction of the cortisol is bioactive, when considering treatments that affect the whole HPA axis, including DHEA, it is essential to measure metabolized cortisol to get a bigger picture.

In urine, we can measure both the total metabolized cortisol (THF) and total metabolized cortisone (THE) excreted throughout the day. These two components better represent the total cortisol production from the adrenal glands than the free cortisol alone. Outside of the HPA axis, metabolism of cortisol occurs with the help of thyroid hormone in the liver. A significant amount of cortisol is also metabolized in adipose tissue.

To best determine total adrenal production of cortisol throughout the day it is important to assess both metabolized cortisol and free cortisol.

When evaluating cortisol levels, it is important to assess the following:

- **The daily pattern of free cortisol throughout the day, looking for low and high levels:**

Abnormal results should be considered along with related symptoms. Remember that with urine results, the "waking" sample reflects the night's total for free cortisol. The sample collected two hours after waking captures the cortisol awakening response, which is typically the time with the most cortisol secretion.

- **The sum of the free cortisol as an expression of the overall tissue cortisol exposure:**

This total of four free cortisol measurements is the best way to assess the total of free cortisol throughout the day, and this result correlates reasonably well to a true 24-hour urine free cortisol. Do be aware that this measurement does not consider transitory shifts in cortisol in the late morning or early afternoon. This number is calculated from the simple addition of the 4 points, so if a single point is very high or very low, it may skew the number up or down especially if it is the morning "B" point, as it is weighted more heavily in the reference range.

- **The total level of cortisol metabolites:**

This total of four free cortisol measurements is the best way to assess the total of free cortisol throughout the day, and this result correlates reasonably well to a true 24-hour urine free cortisol. Do be aware that this measurement does not consider transitory shifts in cortisol in the late morning or early afternoon. This number is calculated from the simple addition of the 4 points, so if a single point is very high or very low, it may skew the number up or down especially if it is the morning "B" point, as it is weighted more heavily in the reference range.

**Free cortisol levels are low. Overall HPA-Axis activity may be somewhat higher than implied by free cortisol as levels of metabolized cortisol are within range. While cortisol production may be lower than needed to meet the body's demands, increased metabolism of cortisol is contributing to the lower levels of free cortisol. Increased clearance of cortisol may be caused by long-term stress, obesity, hyperthyroidism, and other conditions.**

- **A potential preference for cortisol or cortisone (the inactive form):**

Looking at the comparison between the total for free cortisol and free cortisone is NOT the best indication of a person's preference for cortisol or cortisone. The kidney converts cortisol to cortisone in the local tissue. This localized conversion can be seen by comparing cortisol (free) and cortisone levels. To see the patient's preference systemically, it is best to look at which *metabolite* predominates (THF or THE). This preference can be seen in the fan style gauge. This is known as the 11b-HSD index. The enzyme 11b-HSD II converts cortisol to cortisone in the kidneys, saliva gland and colon. 11b-HSD I is more active in the liver, fat cells and the periphery and is responsible for reactivating cortisone to cortisol. Cortisol and cortisone are then metabolized by 5a-reductase to become tetrahydrocortisol (THF) and tetrahydrocortisone (THE) respectively.

## Nutritional Organic Acids

Organic acids are the metabolic byproducts of cellular activity in the body. Organic acid production varies by the individual and can be influenced by foods, environmental toxins, medications or supplements, nutrient status, genetics and more. Organic acids begin to build up when a nutrient cofactor or mineral is not present for a specific reaction to occur. As a response, byproducts (organic acids) build up and can be measured in urine. On the DUTCH test, the organic acids we measure were chosen due to their specific roles in the metabolism and function of enzymes required for hormone and adrenal health and function. As industry standard dictates, the organic acids are measured from the waking sample.

### Methylmalonate (MMA)

Methylmalonic acid is a metabolic byproduct of the Citric Acid Cycle (Krebs cycle). Methylmalonic acid requires adenosylcobalamin for conversion to succinyl-CoA and onto ATP synthesis. If someone does not absorb enough B12 from their diet due to low B12-rich food consumption, low stomach acid, has an autoimmune disorder impacting Intrinsic Factor in the gut (required for B12 absorption), or has a MUT enzyme SNP (required for conversion of MMA to Succinyl coA, dependent on adenosylcobalamin) then MMA will build up. Vitamin B12 is required for COMT activity (estrogen methylation, dopamine breakdown) and PNMT activity (the enzyme that takes norepinephrine to epinephrine), but is also critical for memory, energy production (ATP synthesis), gait and more. When MMA is high, consider supporting B12 through foods, digestive support or supplementation.

### Xanthurenate & Kynurenate

Xanthurenate and kynurenate are metabolic byproducts in the production of tryptophan to NAD in the liver. If either xanthurenate or kynurenate build up in the urine, it can indicate a need for vitamin B6. This need is amplified if BOTH markers are elevated, and often indicates a more severe deficiency of vitamin B6. Vitamin B6 is critical as a co-factor to over 100 important reactions that occur in the human body and is stored in the highest concentration in muscle tissue.

Tryptophan is converted to NAD by the liver and one of the steps in this pathway requires B6. When B6 is insufficient, xanthurenate is made instead. Xanthurenate can also bind to iron and create a complex that increases DNA oxidative damage resulting in higher 8-OHdG levels. If both the xanthurenate and 8OHdG levels are elevated, there is likely an antioxidant insufficiency.

Kynurenate may also become elevated when patients are B6 deficient because of a different, possibly less B6 dependent pathway. While there is always some tryptophan going down the kynurenine pathway towards NAD, and possibly xanthurenate, this process is up regulated by inflammation, estrogen and cortisol elevations. If levels of estrogen or cortisol are high, it may exacerbate kynurenic acid and increase the need for vitamin B6. As the Xanthurenate and Kynurenate pathways lead to biomarkers with other influence in the body, elevations in these markers may not always agree.

**Xanthurenate and Kynurenate are both elevated in this case, so a vitamin B6 deficiency is likely and may be somewhat significant (since both markers are elevated). It is advisable to consider increasing vitamin B6 intake and to be aware of those things listed above that may induce a vitamin B6 deficiency.**

### Pyroglutamate

Pyroglutamate is an intermediate in glutathione recycling and production. Glutathione requires the amino acids cysteine, glycine and glutamate for production. If the body cannot convert pyroglutamate forward to glutathione, it will show up elevated in the urine. High pyroglutamate is an established marker for glutathione deficiency.

Remember that glutathione is one of the most potent antioxidants in the human body and is especially important in getting rid of toxins including the reactive quinone species formed by 4-OH-E1 and 4-OH-E2. This reactive species can damage DNA if not detoxified by either methylation or glutathione.

Some have reported that low pyroglutamate may also be indicative of a need for glutathione; however, this is not established in the scientific literature.

*Note: Pyroglutamate in the urine can also be elevated with Italian cheese consumption. Italian Cheeses (parmesan, etc.) may transiently increase pyroglutamate because they use a thermophilic lactobacilli to ripen the cheese- which our gut breaks down into pyroglutamate. This is not clinically significant and only reflects that they ate this style of cheese (if applicable).*

### Neurotransmitter Metabolites

Neurotransmitters are chemical signals produced by neurons in tissues throughout the body that act as chemical messengers that influence mood, cortisol, heart rate, appetite, muscle contraction, sleep and more. Measuring neurotransmitters directly is difficult because of their instability, and their direct urinary measurements are controversial with respect to how well they reflect the body's level of these neuro-hormones.

Each of the neurotransmitters assessed on the DUTCH test (dopamine, norepinephrine/epinephrine) can be assessed indirectly by measuring their urine metabolites (HVA and VMA respectively). While these metabolites are not a perfect reflection of what is going on in the brain, the scientific literature does affirm their use for a good representation of overall levels of these neurotransmitters in the body.

### **Homovanillate (HVA)**

Homovanillate (HVA) is the primary metabolite of dopamine, a brain and adrenal neurotransmitter that comes from tyrosine (with BH4 and iron as co-factors). Dopamine goes on to create norepinephrine and epinephrine (adrenaline).

Low levels of dopamine are associated with depression, addictions, cravings, apathy, pleasure seeking behaviors, increased sleepiness, impulsivity, tremors, low motivation fatigue and low mood.

High levels of dopamine are associated with agitation, insomnia, mania, hyperactivity, hyper-focus, high stress, anxiety and addictions/cravings/pleasure seeking (to maintain high levels).

### **Vanilmandelate (VMA)**

Vanilmandelate (VMA) is the primary metabolite of norepinephrine and epinephrine (adrenaline). The adrenal gland makes cortisol and DHEA (from the adrenal cortex) as well as norepinephrine and epinephrine (from the adrenal medulla). When adrenal hormone output is low, VMA levels may be low. If HVA levels are significantly higher than VMA, there may be a conversion problem from dopamine to norepinephrine. This case can be caused by a copper or vitamin C deficiency.

The enzymes COMT (methylation of catechols) and MAO are needed to make HVA and VMA from dopamine and norepinephrine respectively. If these enzymes are not working properly, HVA and/or VMA may be low in urine, when circulating levels of dopamine and/or norepinephrine/epinephrine may not be low.

Low levels of norepinephrine/epinephrine are associated with addictions, cravings, fatigue, low blood pressure, low muscle tone, intolerance to exercise, depression, and loss of alertness.

High levels of norepinephrine and epinephrine are associated with feelings of stress, aggression, violence, impatience, anxiety, panic, excess worry/hypervigilance, insomnia, paranoia, increasing tingling/burning, loss of memory, pain sensitivity, high blood pressure and heart palpitations.

### **Melatonin (measured as 6-OHMS)**

Melatonin is considered one of our sleep hormones. It is made predominately by the pineal gland in response to darkness and is stimulated by melanocyte stimulating hormone (MSH). A low MSH is associated with insomnia and an increased perception of pain. Mold exposure can inhibit MSH as well. The majority of our melatonin production comes from the pineal gland, but melatonin is also made in the gut, and to a lesser extent in the bone marrow, lymphocytes, epithelial cells and mast cells.

*The DUTCH test uses the waking (A) sample to test melatonin. The urine sample given on waking reflects overnight hormone production and metabolism. This sample can be used to assess melatonin throughout the night. When patients take a middle of the night sample, both the middle of the night and waking samples are tested and the highest number in ng/mg creatinine is reported.*

### **8-OHdG (8-Hydroxy-2-deoxyguanosine)**

8-OHdG (8-Hydroxy-2-deoxyguanosine) is a marker for estimating DNA damage due to oxidative stress (from ROS creation). 8-OHdG is considered pro-mutagenic and is a biomarker for various cancer and degenerative disease initiation and promotion states. It can be increased by chronic inflammation, increased cell turnover, chronic stress, hypertension, hyperglycemia/pre-diabetes/diabetes, kidney disease, IBD, chronic skin conditions (psoriasis/eczema), depression, atherosclerosis, chronic liver disease, Parkinson's (increasing levels with worsening stages), Diabetic neuropathy, COPD, bladder cancer, or insomnia (to name a few). Studies have shown higher levels in patients with breast and prostate cancers. When levels are elevated it may be prudent to eliminate or reduce any causes and increase the consumption of antioxidant containing foods and/or supplements.

### **Urine Hormone Testing - General Information**

What is actually measured in urine? In blood, most hormones are bound to binding proteins. A small fraction of the total hormone levels are "free" and unbound such that they are active hormones. These free hormones are not found readily in urine except for cortisol and cortisone (because they are much more water soluble than, for



example, testosterone). As such, free cortisol and cortisone can be measured in urine and it is this measurement that nearly all urinary cortisol research is based upon. In the DUTCH Adrenal Profile the diurnal patterns of free cortisol and cortisone are measured by LC-MS/MS.

All other hormones measured (cortisol metabolites, DHEA, and all sex hormones) are excreted in urine predominately after the addition of a glucuronide or sulfate group (to increase water solubility for excretion). As an example, Tajic (Natural Sciences, 1968 publication) found that of the testosterone found in urine, 57-80% was testosterone-glucuronide, 14-42% was testosterone-sulfate, and negligible amounts (<1% for most) was free testosterone. The most likely source of free sex hormones in urine is from contamination from hormonal supplements. To eliminate this potential, we remove free hormones from conjugates (our testing can be used even if vaginal hormones have been given). The glucuronides and sulfates are then broken off of the parent hormones, and the measurement is made. These measurements reflect the bioavailable amount of hormone in most cases as it is only the free, nonprotein-bound fraction in blood/tissue that is available for phase II metabolism (glucuronidation and sulfation) and subsequent urine excretion.

Disclaimer: the filter paper used for sample collection is designed for blood collection, so it is technically considered "research only" for urine collection. Its proper use for urine collection has been thoroughly validated.

## Reference Range Determination (last updated 12.20.2018)

We aim to make the reference ranges for our DUTCH tests as clinically appropriate and useful as possible. This includes the testing of thousands of healthy individuals and combing through the data to exclude those that are not considered "healthy" or "normal" with respect to a particular hormone. As an example, we only use a premenopausal woman's data for estrogen range determination if the associated progesterone result is within the luteal range (days 19-21 when progesterone should be at its peak). We exclude women on birth control or with any conditions that may be related to estrogen production. Over time the database of results for reference ranges has grown quite large. This has allowed us to refine some of the ranges to optimize for clinical utility. The manner in which a metabolite's range is determined can be different depending on the nature of the metabolite. For example, it would not make clinical sense to tell a patient they are deficient in the carcinogenic estrogen metabolite, 4-OH-E1 therefore the lower range limit for this metabolite is set to zero for both men and women. Modestly elevated testosterone is associated with unwanted symptoms in women more so than in men, so the high range limit is set at the 80th percentile in women and the 90th percentile for men. Note: the 90th percentile is defined as a result higher than 90% (9 out of 10) of a healthy population.

Classic reference ranges for disease determination are usually calculated by determining the average value and adding and subtracting two standard deviations from the average, which defines 95% of the population as being "normal." When testing cortisol, for example, these types of two standard deviation ranges are effective for determining if a patient might have Addison's (very low cortisol) or Cushing's (very high cortisol) Disease. Our ranges are set more tightly to be optimally used for Functional Medicine practices.

Below you will find a description of the range for each test:

Female Reference Ranges (Updated 08.21.2019)									
	Low%	High%	Low	High		Low%	High%	Low	High
b-Pregnanediol	20%	90%	600	2000	Cortisol A (waking)	20%	90%	10	50
a-Pregnanediol	20%	90%	200	740	Cortisol B (morning)	20%	90%	30	130
Estrone (E1)	20%	80%	12	26	Cortisol C (~5pm)	20%	90%	7	30
Estradiol (E2)	20%	80%	1.8	4.5	Cortisol D (bed)	0	90%	0	14
Estriol (E3)	20%	80%	5	18	Cortisone A (waking)	20%	90%	40	120
2-OH-E1	20%	80%	5.1	13.1	Cortisone B (morning)	20%	90%	90	230
4-OH-E1	0	80%	0	1.8	Cortisone C (~5pm)	20%	90%	32	110
16-OH-E1	20%	80%	0.7	2.6	Cortisone D (bed)	0	90%	0	55
2-Methoxy-E1	20%	80%	2.5	6.5	Melatonin (6-OHMS)	20%	90%	10	85
2-OH-E2	0	80%	0	1.2	8-OHdG	0	90%	0	5.2
4-OH-E2	20%	80%	0	0.5	Methylmalonate	0	90%	0	2.2
2-Methoxy-E2	20%	80%	0	0.7	Xanthurenate	0	90%	0	1.4
DHEA-S	20%	90%	20	750	Kynurenate	0	90%	0	7.3
Androsterone	20%	80%	200	1650	Pyroglutamate	10%	90%	32	60
Etiocolanalone	20%	80%	200	1000	Homovanillate	10%	95%	4	13
Testosterone	20%	80%	2.3	14	Vanilmandelate	10%	95%	2.4	6.4
5a-DHT	20%	80%	0	6.6					
5a-Androstenediol	20%	80%	12	30	<b>Calculated Values</b>				
5b-Androstenediol	20%	80%	20	75	Total DHEA Production	20%	80%	500	3000
Epi-Testosterone	20%	80%	2.3	14	Total Estrogens	20%	80%	35	70
a-THF	20%	90%	75	370	Metabolized Cortisol	20%	90%	2750	6500
b-THF	20%	90%	1050	2500	24hr Free Cortisol	20%	90%	65	200
b-THE	20%	90%	1550	3800	24hr Free Cortisone	20%	90%	220	450

*% = population percentile: Example - a high limit of 90% means results higher than 90% of the women tested for the reference range will be designated as "high."*

### Provider Notes:

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